

SYBR Green qPCR Service

Customer Information

Customer Name:

Institute: College of Veterinary Science

Telephone:

Address:

Email:

1. Samples:

Item	Sample Name	qPCR Sample code
1	Control	HB211
2	Acute Quinalphos toxicity(ACT)	HB212
3	Chronic Quinalphos toxicity(CQT)	HB213
4	Acute Dimethoate Toxicity(ADT)	HB214
5	Chronoc Dimethoate Toxicity(CDT)	HB215
6		
7		
8		

Comparisons:

	Treatment	Control
.....		

2. A. Primers

This qPCR service request included 8 target genes and 10 samples.

Sample Name	Primer Name
Control	AvBD1
Acute Quinalphos toxicity(ACT)	AvBD6
Chronic Quinalphos toxicity(CQT)	AvBD7
Acute Dimethoate Toxicity(ADT)	GAPDH
Chronoc Dimethoate Toxicity(CDT)	ACTB

B. Procedure

Protocol for DNA Isolation

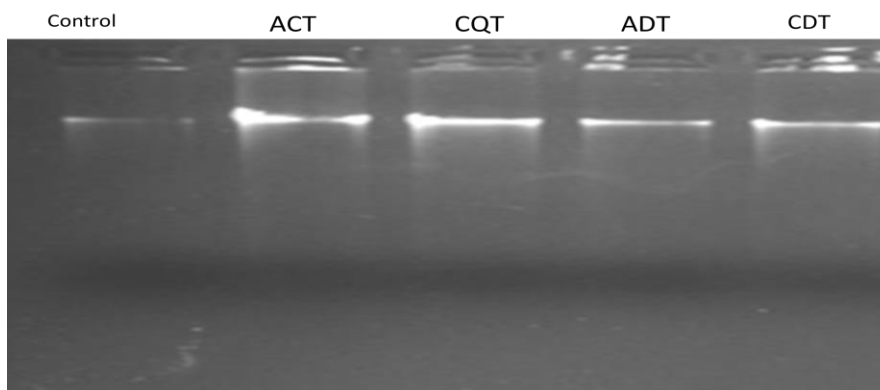
1. Cut up to 50-100 mg tissue into small pieces, and place in a 2.0 ml micro centrifuge tube. Add 350 μ l Buffer TTL.
2. Add 20 μ l Proteinase K and 5 μ l RNase A. Mix thoroughly by vortexing, and incubate at 65oC until the tissue is completely lysed. Briefly invert the tube during incubation.

3. Add 300 µl Buffer TBL to the sample, Vortex for 15-20 seconds and incubate at 65°C for 10 min and mix thoroughly by inverting. Then add 150 µl ethanol (96-100%), mixed by inverting a tube 15-20 times or Pulse vortexing. Incubate for 1 min at room temperature (18-25°C). Briefly centrifuge the MCT (1-2 sec at 2,000 x g) to remove drops from the lid (short spin only).
4. Transfer 650 µl lysate to the TransPure™ spin column, and centrifuge at 10,000rpm for 2 min. discard the flow through liquid.
5. Repeat above step, until the entire sample has been processed and retain column for further processing.
6. Place the column into same collection tube. Add 500 µl of Buffer TW. Centrifuge at 10,000rpm for 1 min. Discard the flow through.
7. Place the column into same collection tube. Add 650 µl of TW I. Centrifuge at 10,000rpm for 1 min. Discard the flow through.
8. Repeat the above “step-7” again.
9. Place the empty TransPure™ spin column, with the lid open into the same collection tube and centrifuge at 10,000 rpm for 2 min.
10. Place the column into a new sterile 1.5 ml Eppendorf tube, add 30 µl preheated (75°C-80°C) Elution buffer. Incubate at room temperature for 5 min. (perform this step twice 30 µl + 30 µl= 60 µl)
11. Centrifuge 10,000 rpm for 1 min to elute pure Tissue gDNA. The first elution normally yields 60-70% of DNA bound. A second elution with another 30 µl buffer will yield another 20 % of the DNA.
12. Discard the column, and save elute. Do not reuse binding columns or collection tubes.

Agarose gel electrophoresis

The amplified PCR products were separated by electrophoresis in 0.8% agarose gel run in 1× TAE buffer at 50V for 30 to 45 min. till DNA fragments are migrated well. The gel was photographed on UVP version gel documentation system, stored and analyzed using a gel documentation system with built in software.

Gel Photo:



3. A. Quantitative PCR (q-PCR)

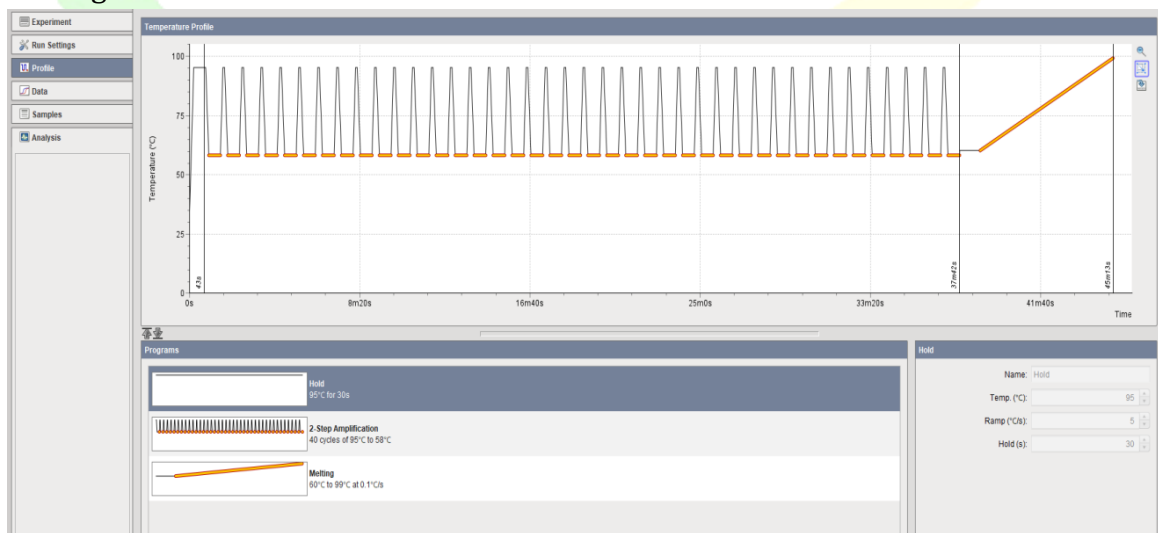
Each sample was tested in duplicate. Each reaction included 20 ng cDNA, 10pocomole of forward and reverse primers, and 2X Fast SYBR Green PCR Master Mix (TB Green Premix Ex Taq II, # RR820A).

10 μ l reaction volumes were used according to the table below:

Fast SYBR master mix(μ l)	nuclease-free water(μ l)	RT product (20 ng/ μ l)	F+R primer (10pM)	total volume (μ l)
5	3	1	1	10

B. Temperature & Program file

A **Tata MD CHECK Express real time PCR** machine was used with the following program: (95°C → 30sec), and 40 cycles (95°C → 5sec, 58°C → 30sec) followed by Melting curve 60°C to 99°C.



4. Data Analysis

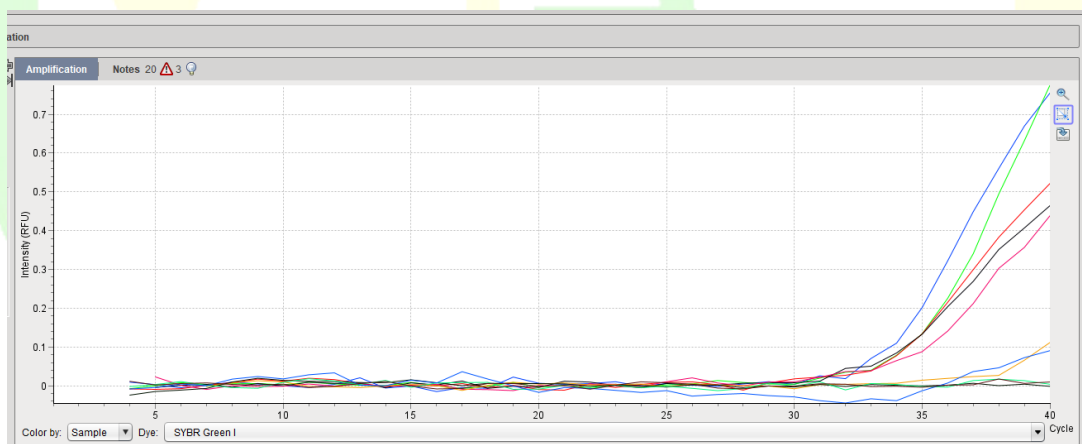
MyGo pro PCR software 3.6 was used for experimental setup and data analysis. Target gene qPCR data were determined by Cq Value.

A. AvBD1Gene

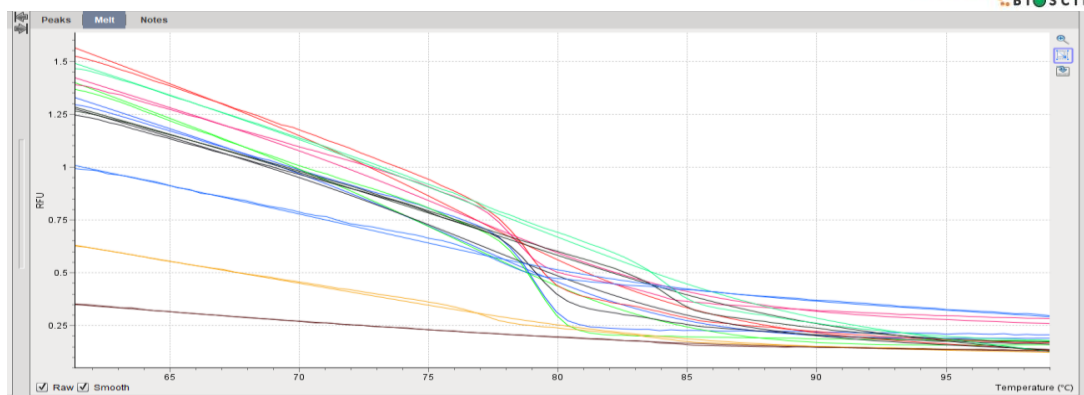
B. Plate Set up Real time PCR

	1	2	3	4	5	6	7	8
A	Control	Control_1	AQT	AQT_1	CQT	CQT_1	ADT	ADT_1
B	CDT	CDT_1						
C								
D								

C. Amplification Curve



D. Melting curve



E.

F. CT VALUE

Sample Name	Primer	CT values
Control	AvBD1	32.63
Control_1	AvBD1	33.972
AQT	AvBD1	32.795
AQT_1	AvBD1	32.635
CQT	AvBD1	33.6
CQT_1	AvBD1	36.749
ADT	AvBD1	
ADT_1	AvBD1	
CDT	AvBD1	
CDT_1	AvBD1	

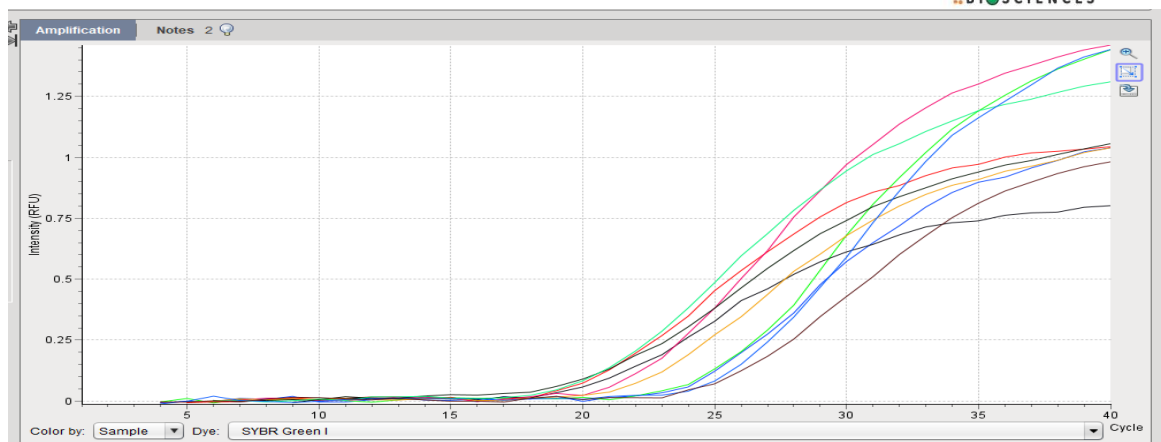
5.

A. AvBD6 Gene

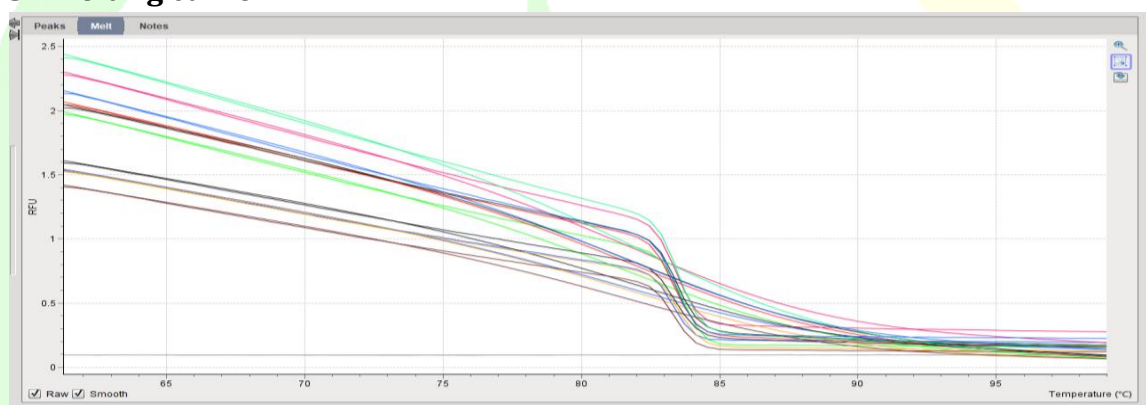
Plate Set up Real time PCR

	1	2	3	4	5	6	7	8
A	Control	Control_1	AQT	AQT_1	CQT	CQT_1	ADT	ADT_1
B	CDT	CDT_1						
C								
D								

B. Amplification Curve



C. Melting curve



D. CT value

Sample Name	Primer	CT values
Control	AvBD6	23.613
Control_1	AvBD6	24.511
AQT	AvBD6	19.427
AQT_1	AvBD6	19.775
CQT	AvBD6	21.358
CQT_1	AvBD6	21.407
ADT	AvBD6	20.336
ADT_1	AvBD6	19.801
CDT	AvBD6	24.897
CDT_1	AvBD6	24.535

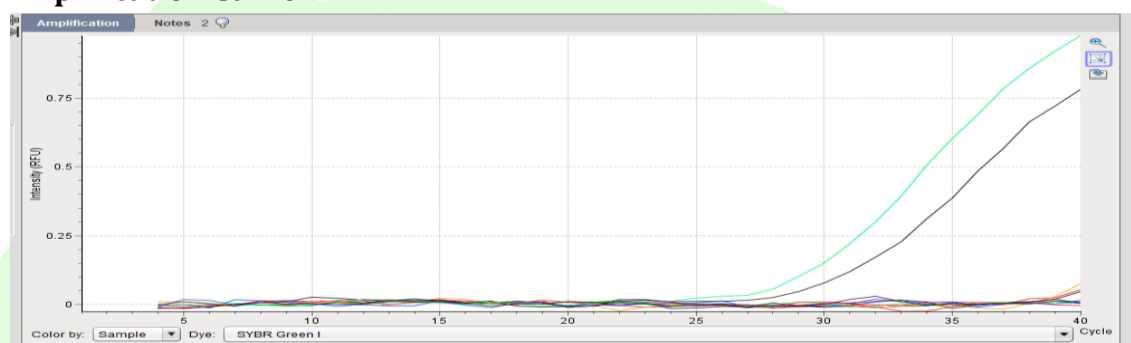
6A. AvBD7 Gene

Plate Set up Real time PCR

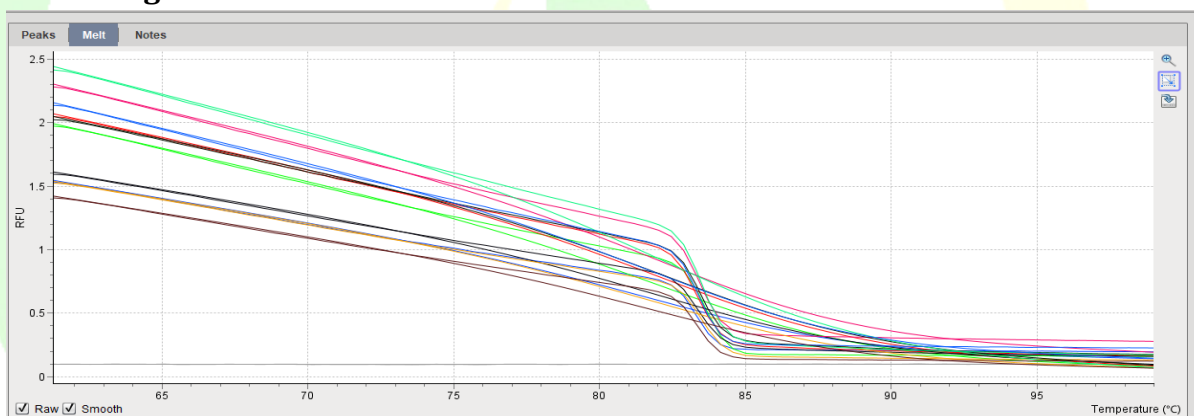
	1	2	3	4	5	6	7	8
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A	Control	Control_1	AQT	AQT_1	CQT	CQT_1	ADT	ADT_1
B	CDT	CDT_1						
C								
D								

E. Amplification Curve



F. Melting curve



G. CT value

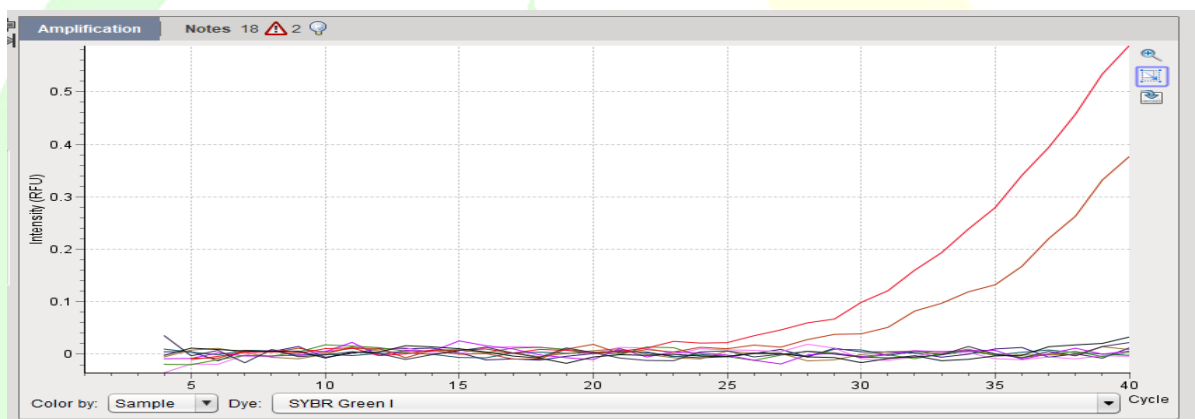
Sample Name	Primer	CT values
Control	AvBD7	
Control_1	AvBD7	
AQT	AvBD7	
AQT_1	AvBD7	
CQT	AvBD7	
CQT_1	AvBD7	
ADT	AvBD7	30.153
ADT_1	AvBD7	28.652
CDT	AvBD7	
CDT_1	AvBD7	

7A. GAPDH Gene

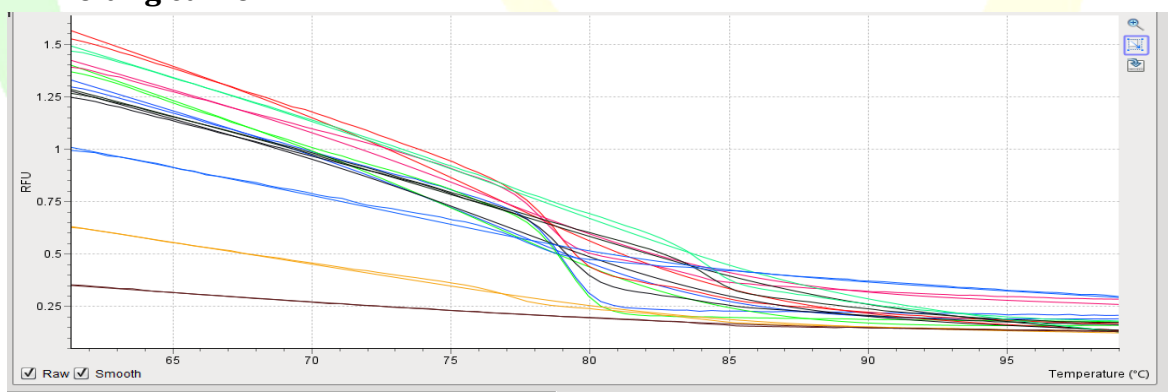
Plate Set up Real time PCR

	1	2	3	4	5	6	7	8
A	Control	Control_1	AQT	AQT_1	CQT	CQT_1	ADT	ADT_1
B	CDT	CDT_1						
C								
D								

H. Amplification Curve



I. Melting curve



J. CT value

Sample Name	Primer	CT values
Control	GAPDH	
Control_1	GAPDH	
AQT	GAPDH	
AQT_1	GAPDH	
CQT	GAPDH	

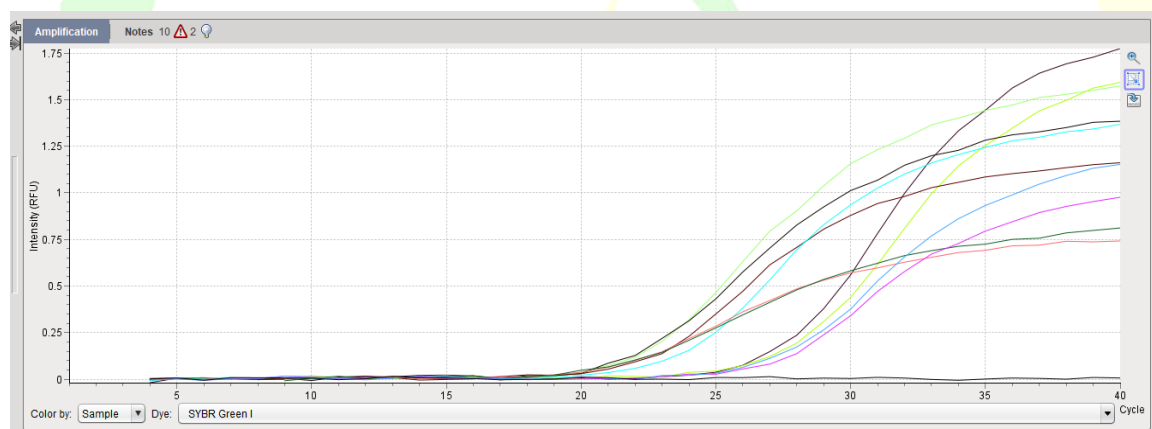
CQT_1	GAPDH	
ADT	GAPDH	34.748
ADT_1	GAPDH	33.327
CDT	GAPDH	
CDT_1	GAPDH	

8 A. ACTB Gene

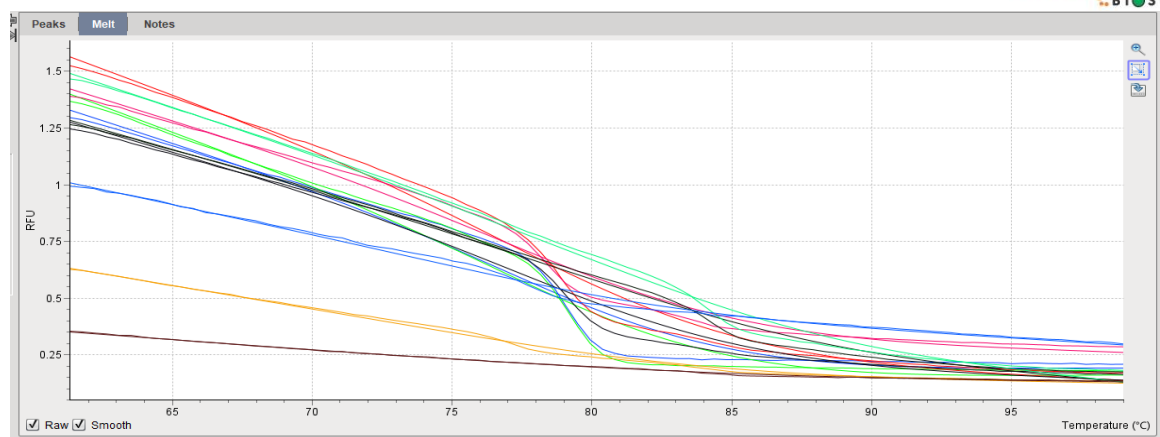
Plate Set up Real time PCR

	1	2	3	4	5	6	7	8
A	Control	Control_1	AQT	AQT_1	CQT	CQT_1	ADT	ADT_1
B	CDT	CDT_1						
C								
D								

K. Amplification Curve



L. Melting curve



M. CT value

Sample Name	Primer	CT values
Control	ACTB	26.636
Control_1	ACTB	27.112
AQT	ACTB	21.385
AQT_1	ACTB	21.063
CQT	ACTB	21.685
CQT_1	ACTB	22.875
ADT	ACTB	20.411
ADT_1	ACTB	20.566
CDT	ACTB	26.38
CDT_1	ACTB	26.512